



Artificial color tuning of firefly luminescence: Theoretical mutation by tuning electrostatic interactions between protein and luciferin

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ABSTRACT

Electrostatic interactions between firefly oxyluciferin and the surrounding proteins were analyzed, and the amino acids important for controlling emission energy were identified. We propose Arg223Ala, Glu344Ala, and Asp422Ala mutations in firefly oxyluciferase of *Photinus pyralis*, which artificially change the luminescence color by tuning the electrostatic effect from the luciferase proteins. In the theoretical mutation simulation, the emission energy of the triple mutant was estimated to be 2.05 eV (602 nm, red-dish-orange), which is 0.18 eV lower than that of the wild type (2.23 eV, 557 nm, yellow-green). For calculating the emission energies, we used the symmetry-adapted cluster-configuration interaction (SAC-CI) method.

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1. Introduction

Firefly luciferase (Luc) is an oxygenase enzyme which catalyzes the light-emitting reaction. Luc oxygenates firefly-luciferin and produces oxyluciferin in the electronically excited state (see Fig. 1a) [1–5]. During the decay process to the ground state, keto-oxyluciferin (oxyLH₂) shows yellow-green emission [1,2]. The quantum yield of the luminescence is known to be very high [6]. Firefly Luc has successfully been applied to various fields of bio-imaging [2] to assay ATP generation [7], gene expression [8], and biosensors for environmental pollutants [8]. It is desirable to develop Luc showing different emission colors [2]. In particular, red-light emitting Luc variants are useful for the applications to mammalian cells which absorb shorter wavelength [2,9].

Recently, a red emitting Ser284Thr mutant (615 nm, 2.02 eV) and a green emitting Val241Ile/Gly246Ala/Phe250Ser triple mutant (549 nm, 2.26 eV) of *Photinus pyralis* (Ppy) have been developed and showed favorable spectroscopic and kinetic properties [10]. A synergistic effect was observed in a mutagenesis study [11], indicating that the active-site local structure and the steric effect were related to the spectral tuning [11]. Regarding *Luciola cruciata*, two red emitting mutants, Ile288Val (613 nm, 2.02 eV) and Ile288Ala (613 nm, 2.02 eV) were also produced [12]. According to the X-ray structural observations, the spectral shift correlates with the size of the side chain in the 288th amino acid [12]. These successful examples are thus based on the steric constraint which

controls the structural relaxation of the excited oxyLH₂ before the emission (Fig. 1b).

In our previous study [13], we analyzed the emission color tuning mechanism of Ppy Luc, why red-light in chemiluminescence and yellow-green in Luc. The oxyLH₂ structural distortion induced in the Luc environment reduces the emission energy by about 0.24–0.39 eV. The C–C dihedral angles between the two rings were ascribed as the main structural difference. This result indicated that the emission energies might be controlled by modifying the structural relaxations in the excited state of oxyLH₂, which supports the steric constraint strategy [10–12].

We have also shown that the quantum and classical Coulombic interactions between oxyLH₂ and Luc increase the emission energy by about 0.50–0.60 eV. We performed quantum-theoretical calculations and clarified the crucial role of the protein-electrostatic effects on the emission energy. Since the charge distribution of oxyLH₂ changes upon the electronic transition, the emission energy is sensitive to the protein-electrostatic environment as schematically shown in Fig. 1c. This effect was shown to be a major factor for making the emission color to be yellow-green [13]. Therefore, it is a reasonable strategy to control the electrostatic interactions between oxyLH₂ and Luc proteins. Once the interactions between oxyLH₂ and amino acids are analyzed, we are able to identify several key amino acids which control the emission energy. It is therefore possible to propose new mutation experiments for the emission-spectral tuning.

For calculating emission energies, we used the symmetry-adapted cluster-configuration interaction (SAC-CI) method [14–17]. The method has been applied to more than 150 molecules and established as an accurate electronic-structure theory for the ground and excited states [18–20]. In our recent study, we

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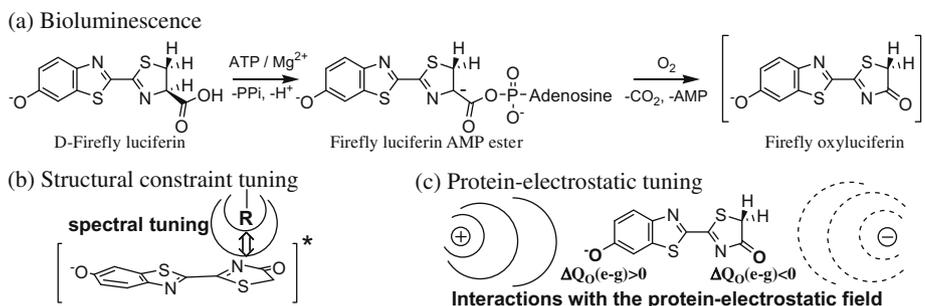


Fig. 1. (a) Reaction scheme of the firefly-luciferin enzyme reaction. (b) Protein-electrostatic effect on the excited state of oxyLH_2 . $\Delta Q_{\text{O}}(\text{e-g})$ is the atomic-charge difference of oxygen atoms between the excited and ground states of oxyLH_2 ($\Delta Q_{\text{O}}(\text{e-g}) = Q_{\text{O}}(\text{e}) - Q_{\text{O}}(\text{g})$).

successfully reproduced the emission energies of oxyLH_2 observed in chemiluminescence and bioluminescence [13]. The method also successfully applied to analyze the color tuning mechanisms in retinal proteins [21–24] and green-fluorescent proteins [25,26].

The aim of the present study is to propose mutations to control the electrostatic interactions between oxyLH_2 and Luc for tuning the color of the firefly luminescence. First, the results of the decomposition analysis of the electrostatic interaction between oxyLH_2 and the surrounding molecules are described. Next, on the basis of the electrostatic strategy, our new proposals of the mutations are proposed. Finally, to numerically examine our proposals, we performed the SAC-CI emission energy calculations, and the results of the theoretical mutation simulations are described.

2. Computational details

The X-ray structure (1LCI [27]) and the working model proposed in the experimental studies [28–32] were used for constructing the oxyLH_2 -Luc binding complexes. Molecular dynamics calculations with AMBER 94 force-field [33] were performed to find the stable structures. The force-field parameters for oxyLH_2 were designed for the first excited state [13]. When all of the atoms in an amino acid were located more than 7.0 Å away from oxyLH_2 , the atomic coordinates were frozen in the molecular dynamics (MD) calculations. We took three stable structures from the trajectory. These three had different AMP conformations. There were only minor differences in the other part of the structures. Starting from these three, we raised the temperature stepwise up to 300 K, in which total 60 ps MD simulations were performed. Then we performed 30 ps MD simulations at 300 K. After the MD calculations, the molecular mechanics (MM) energy minimizations were performed. The MM minimized structures were used for the initial guesses for the configuration interaction singles (CIS) geometry optimizations.

In the CIS calculations, oxyLH_2 , AMP, and amino acids in the first solvation layer (ARG218, HIS245, PHE247, GLY341, LEU342, SER347, ALA348, and the crystal water HOH45) were treated with the quantum mechanical (QM) method. The rest of the protein effect was included by the classical point-charge model defined by the MM parameters. Geometry of oxyLH_2 was optimized in the CIS calculations. The other atoms in the QM region were fixed to the MM optimized structures. The basis sets of 6-31g* were used for oxyLH_2 , and the 6-31g set for the rest of the QM atoms.

Single-point SAC-CI calculations were performed at the optimized geometries to evaluate the fluorescence energies. oxyLH_2 , His245, Arg218, and the phosphate group of AMP were treated quantum mechanically, and the rest was done by a classical point-charge model. All valence orbitals were included in the active space of the SAC-CI calculations. Perturbation selection was performed to select the double excitation operators [34]. The en-

ergy thresholds of 5.0×10^{-6} and 5.0×10^{-7} hartree (LevelTwo) were used for the ground and excited states, respectively. For the basis sets, we used Huzinaga–Dunning double-zeta basis sets [35] augmented with the single-polarization d-function [35]. The calculated emission energy for the wild type, 2.23 eV, was in good agreement with the experimental one, 2.33 eV [13].

In constructing the computational model, we trusted the X-ray structure. Comparing the structures available in the Protein Data Bank [27,36], the luciferase structures were not so sensitive to the inhibitor binding in the active-site [36]. In our previous study [13], including the oxyluciferin in the binding site gave only small structural changes. One reason is that luciferase has open space in the binding site. If there is no inhibitor inside, the space is almost filled by water molecules.

The QM and MM calculations were performed using the GAUSSIAN 03 [37] and TINKER programs [38–40], respectively.

3. Results and discussions

3.1. Role of the protein– oxyLH_2 interactions behind the yellow–green emission of the wild type

Since bioluminescences originate from the electronic transition, the charge distribution of the chromophore changes after the luminescence. In our previous study, the emission color tuning mechanism was ascribed to the increase/decrease in the charges of the carbonyl O atoms [13]. However, the transition is better characterized as an intramolecular charge-transfer from the benzothiazole ring to the thiazole moiety. Fig. 2 shows the orbital distributions of the lowest unoccupied molecular orbital (LUMO) and the highest occupied molecular orbital (HOMO). As seen in Fig. 2b, the benzothiazole ring in the HOMO is more populated than the thiazole ring. In contrast, the LUMO shown in Fig. 2a distributes slightly

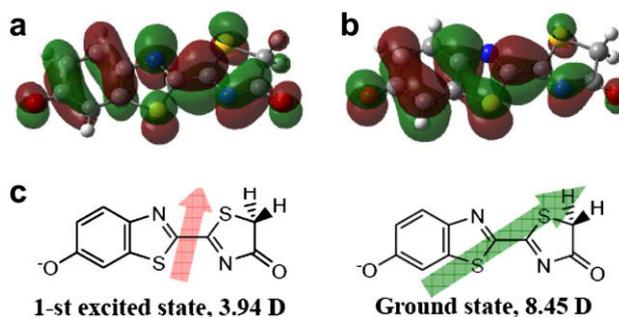


Fig. 2. Orbital distributions of (a) LUMO and (b) HOMO. (c) Dipole moment of oxyLH_2 in the first excited and ground states.

more in the thiazole moiety. Dipole moments clearly reflect the charge-transfer character as shown in Fig. 2c. OxyLH₂ is polar in the ground state (8.45 D), but, in the excited state, the dipole moment is much smaller (3.94 D).

Due to the charge-transfer character, the emission energy of oxyLH₂ is influenced by the electrostatic field generated by the Luc environment, which were the major origin of the yellow-green emission in the wild type (WT). We analyzed the protein-electrostatic effect at the amino acids resolution as follows.

$$\begin{aligned} \Delta E &= \sum_M^{\text{Amino acids}} \Delta E_M = \sum_M^{\text{Amino acids}} \{E_M^{\text{Ex}} - E_M^{\text{G}}\} \\ &= \sum_M^{\text{Amino acids}} \left\{ \sum_{B \in M} \sum_{A \in \text{OxyLH}_2} \frac{(Q_A^{\text{Ex}} - Q_A^{\text{G}}) Q_B}{r_{AB}} \right\} \end{aligned} \quad (1)$$

The ΔE is the difference in the interaction energy between the ground and excited states, and can be easily decomposed into the contribution from each amino acid, $\Delta E_M (= E_M^{\text{Ex}} - E_M^{\text{G}})$. The E_M^{Ex} and E_M^{G} are the electrostatic interaction energies in the excited and ground states, respectively. We evaluated the interaction energy as the classical Coulombic interaction between oxyLH₂ and the surrounding molecules. The atomic-charges of oxyLH₂ in the ground and the excited states, Q_A^{G} and Q_A^{Ex} , were obtained by the electrostatic potential fitting using HF and CIS wave functions, respectively. The Q_B for the atoms in Luc were taken from the AMBER 94 force-field [33].

The result of the decomposition analysis is shown in Fig. 3. The blue- and red-shift contributions in the emission energy caused by each amino acid were indicated by the positive and negative regions of the diagram, respectively. It is clearly shown that Arg218, His245, and AMP (phosphate group) give the primary contributions. Arg218 and AMP cause the blue-shifts of 0.29 and 0.21 eV, respectively, and His245 contributes to the red-shift of 0.17 eV [13]. In a previous experimental mutation study, fluorescence energies of Arg218Ala and Arg218Gln mutants were observed to be 2.03 (611) and 2.04 eV (608 nm), respectively, which were around 0.2 eV smaller than that of the wild type [32]. These experimental results qualitatively agree to our analysis, because the positively-charged residue at the 218th position causes the blue shift. A Ser347Ala mutation red-shift around 0.17 eV [30] also supports the present theoretical result.

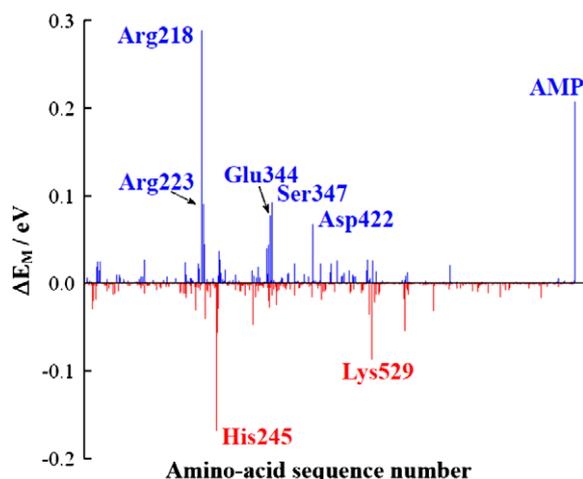


Fig. 3. Decomposition analysis of $\Delta E (= E^{\text{Ex}} - E^{\text{G}})$, difference of the interaction energies in the ground and the excited states). ΔE was decomposed into contributions from the protein residues and AMP (ΔE_M). Positive and negative contributions indicate spectral blue- and red-shifts in fluorescence, respectively.

In this study, we focus on the secondary important contributions from the amino acids relatively distant from the binding site: blue-shift contributions from Arg223, Glu344, and Asp422, and red-shift contribution from Lys529 as shown in Fig. 3. The positions and structures of these amino acids were shown in Fig. 4. Since Arg223 is positively-charged and located close to Arg218, this amino acid contributes the positive electrostatic potential as shown in Fig. 1c. In contrast, negatively charged Glu344 and Asp422 provide the negative electrostatic potential in the same way as the AMP phosphate group.

3.2. Artificial red- and yellow-emitting firefly luciferases: theoretical mutation simulations

On the basis of the analysis described above, we point out some amino acids to obtain red-emitting firefly luciferases. As shown in Fig. 3, Arg223, Glu344, and Asp422 have blue-shift contributions in the emission energies of the wild type. When they are replaced by a neutral residue as alanine, the blue-shift effect should disappear, and the emission energy is expected to show red-shifts. Since the distances between oxyLH₂ and the target residues are around 7–12 Å, we assumed the structural rearrangements around the oxyLH₂ binding site causes minor effect. On the other hand, Arg218, His245, and Ser347 were excluded. Since these residues are in direct contact with oxyLH₂, the mutations would change the oxyLH₂ binding conformations. Once the primary binding site residue was lost, it would be very difficult to find a reasonable binding structure between oxyLH₂ and Luc in computational point of view. The experimental mutations at Arg218, His245, and Ser347 were already reported [29,30,32]. Significant spectral shifts were observed in these mutants [32], which qualitatively agree to our result of the electrostatic analysis. However, since the mutation, especially Arg218, also dropped the oxyLH₂ binding capability of the luciferase, the oxygenase catalytic reactions became less efficient than that in the wild type [29,30,32].

Next we performed theoretical mutation simulations. The Arg223, Glu344, and Asp422 were replaced by alanines, and the structures of the amino acids were optimized by the MM calculations. The SAC-Cl calculations were performed to evaluate the fluorescence energies of the mutants.

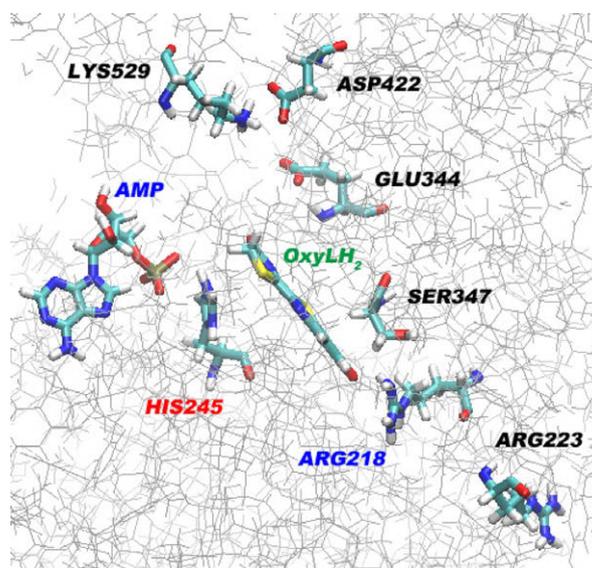


Fig. 4. Amino acids relevant to the emission color tuning. The structure of the Luc-oxyLH₂ complex was optimized for the first excited state of oxyLH₂.

Table 1
Emission energies of the mutants calculated by the SAC-CI method.

System	Theoretical			Exptl. ^b
	SAC-CI	Spectral shift	Shifted ^{a,b}	
Wild type	2.33 ^c	–	2.23 (557)	2.23 (557) ^d
R223A	2.27	–0.06	2.17 (571)	
E344A	2.22	–0.11	2.12 (585)	
D422A	2.25	–0.08	2.15 (577)	
R223A/E344A/D422A	2.15	–0.18	2.05 (605)	

^a Emission energies were shifted by –0.1 eV, so that the emission energy of the wild type became equal to the experimental value.

^b Unit is in eV. The numbers in parentheses are in nm unit.

^c Ref. [13].

^d Ref. [10].

In Table 1, the calculated emission energies were summarized. In the Arg223Ala, Glu344Ala, and Asp422Ala mutants, the emission energies were calculated to be 2.27 eV, 2.22 eV, and 2.25 eV, respectively. The estimated spectral red-shifts are 0.06 eV, 0.11 eV, and 0.08 eV, respectively. The amounts of the shifts qualitatively agree with those obtained by the analysis as shown in Fig. 3. Table 1 also showed the ‘Shifted’ emission energies which were shifted by –0.10 eV so that the theoretical result of the wild type became equal to the experimental one. The expected emission energies of the Arg223Ala, Glu344Ala, and Asp422Ala mutants were 2.17 eV (571 nm), 2.12 eV (585 nm), and 2.15 eV (577 nm), respectively. Although the amounts of the shifts are not so significant, we expect yellow emissions from these single mutants.

We further examined a triple mutant, Arg223Ala/Glu344Ala/Asp422Ala. The amount of the estimated red-shift became 0.18 eV and the expected emission energy became 2.05 eV (605 nm), which is the reddish-orange emission.

To the best of our knowledge, none of the amino acids pointed out in the present study has been tested in previous experimental mutations. Since the targeted amino acids locate relatively far from oxylH₂ binding site, the mutations would not directly change the binding site. Therefore, we expect that the mutation effects proposed based on the present and previous studies [10–12] would provide additive effects in controlling the emission energy. We strongly hope that the present theoretical predictions are tested by real mutation experiments.

4. Conclusions

There are several successful firefly Luc mutants showing the red and green emissions [10,12]. The mechanism proposed in these studies was based on the steric constraint [11,12]. On the other hand, we found that the protein-electrostatic effect gave the largest contribution to the blue-shift in the emission energy [13]. In this study, we proposed three amino acids for the mutation experiments, Arg223, Glu344, and Asp422 in *Ppy* Luc.

Theoretical mutation simulations using the SAC-CI method showed that the Arg223Ala, Glu344Ala, and Asp422Ala mutations showed spectral red-shift of 0.06, 0.11, and 0.08 eV, respectively. A triple mutant, Arg223Ala/Glu344Ala/Asp422Ala, showed a larger red-shift of 0.18 eV. The emission energy expected is 2.05 eV (605 nm, reddish-orange). We thus wish that our proposal would add color variations in addition to the existing successful mutants [10,12].

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